

Conserved synteny at the protein family level reveals genes underlying *Shewanella* species' cold tolerance and predicts their novel phenotypes

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Received: 28 May 2009 / Revised: 24 August 2009 / Accepted: 10 September 2009 / Published online: 3 October 2009
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Abstract Bacteria of the genus *Shewanella* can thrive in different environments and demonstrate significant variability in their metabolic and ecophysiological capabilities including cold and salt tolerance. Genomic characteristics underlying this variability across species are largely unknown. In this study, we address the problem by a comparison of the physiological, metabolic, and genomic characteristics of 19 sequenced *Shewanella* species. We have employed two novel approaches based on association of a phenotypic trait with the number of the trait-specific

protein families (Pfam domains) and on the conservation of synteny (order in the genome) of the trait-related genes. Our first approach is top-down and involves experimental evaluation and quantification of the species' cold tolerance followed by identification of the correlated Pfam domains and genes with a conserved synteny. The second, a bottom-up approach, predicts novel phenotypes of the species by calculating profiles of each Pfam domain among their genomes and following pair-wise correlation of the profiles and their network clustering. Using the first approach, we find a link between cold and salt tolerance of the species and the presence in the genome of a Na^+/H^+ antiporter gene cluster. Other cold-tolerance-related genes include peptidases, chemotaxis sensory transducer proteins, a cysteine exporter, and helicases. Using the bottom-up approach, we found several novel phenotypes in the newly sequenced *Shewanella* species, including degradation of aromatic compounds by an aerobic hybrid pathway in *Shewanella woodyi*, degradation of ethanolamine by *Shewanella benthica*, and propanediol degradation by *Shewanella putrefaciens* CN32 and *Shewanella* sp. W3-18-1.

Electronic supplementary material The online version of this article (doi:10.1007/s10142-009-0142-y) contains supplementary material, which is available to authorized users.

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Keywords Phenotypic trait · Bacteria · Molecular mechanisms of cold tolerance · *Shewanella* · Protein families

Introduction

Shewanella species are known for their ability to utilize a diverse set of electron acceptors for anaerobic respiration. This capability makes the bacterium important for remediation of environments contaminated with nitroaromatic compounds

chromium, plutonium, uranium, and other radionuclides (Gralnick and Hau 2006; Fredrickson et al. 2008). Members of the genus typically inhabit chemically stratified sedimentary and aquatic systems with differences in available sources of nutrients and electron acceptors as well as in physical characteristics such as temperature, salinity, pH, and pressure (Gralnick and Hau 2006). To live in these diverse environments and to compete with other microorganisms, the *Shewanellae* have evolved different physiological and metabolic characteristics that are fine-tuned to their environmental niches. At present, 19 *Shewanella* strains from diverse environments have been sequenced providing a unique opportunity for comprehensive comparative analyses of their genomes and for revealing genomic determinants of the various phenotypes of the species. Objectives of the study were (1) to link complex ecophysiological traits of *Shewanella* species, such as cold and salt tolerance, to specific genes and molecular mechanisms, and (2) to reveal phenotypic and metabolic capabilities of the newly sequenced species from their genomic characteristics.

Current computational technologies are limited in their ability to associate a specific phenotypic characteristic of an organism with its genomic signature in a comprehensive manner. Traditional linkage analysis (Lander and Kruglyak 1995) allows mapping of phenotypic differences between individuals to their genotypic differences but the analysis was applied to eukaryotes and investigated intra-individual genetic differences within the same species. Other known approaches include phylogenetic profiling based on correlations between the presence/absence of phenotypes and the presence/absence of genomic characteristics (e.g., specific enzymes, Pfam domains, clusters of orthologous groups of proteins (COGs), or orthologous genes) in a set of species (Venkateswaran et al. 1999; Liu et al. 2006; Ranea et al. 2007; Date and Peregrin-Alvarez 2008). More flexible, although less-specific, strategies are phylogenetic pattern analysis (Makarova et al. 2003) and natural language processing approaches (Korbel et al. 2005), methods used to find associations between COGs and phenotypes. To our knowledge, none of the current computational methods that address genotype and phenotype associations compare genomes in terms of synteny (a genomic locus with a conserved order of similar genes) at the protein family level calculating the number of each protein family instead of its binary characteristic (present/absent) and correlating this number with a quantitative outcome of a complex phenotype. Feasibility of the approach stems from knowledge about molecular mechanisms involved in development of new phenotypic traits in living organisms (Fig. 1S). It is believed that the enormous interspecies phenotypic variation in both eukaryotes and prokaryotes is caused by elaboration on the existing gene structures rather than by *de novo* invention of genes. For example, combining existing

domains in novel gene architectures, also known as domain shuffling, has been estimated to have involved up to 20% of eukaryotic genes (Long et al. 1995). In addition to domain shuffling, new gene structures may be created by duplication of a gene with subsequent accumulation of mutations (Long et al. 2003) or by acquisition of genes by horizontal transfer, a well-known mechanism in prokaryotes (Gogarten and Townsend 2005). Duplication of protein domains, if they are important for a particular phenotypic trait, in combination with deletion of domains, if they are not needed, leads to an enrichment in the genome of domains specific for phenotypic traits and to a conserved domain synteny in the microorganisms implementing the trait (Apic et al. 2001; Methe et al. 2003; Madera et al. 2004; Orengo and Thornton 2005).

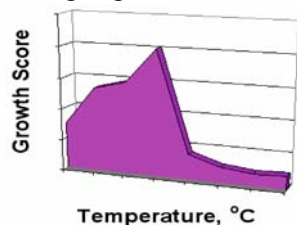
Based on the genomic mechanisms described above, we propose to link genotype and phenotype of the *Shewanella* species using (1) correlation between the number of trait-specific protein domains as defined by the Pfam database (also referred as domain enrichment) and a quantitative characteristic of the phenotypic trait and (2) conserved synteny of the trait-specific genes (Fig. 1). The first, a top-down approach, involves quantification and comparison of a higher-level characteristic, the bacterial cold tolerance, to reveal genomic characteristics and specific genes related to the phenotype (Fig. 1a). The second, a bottom-up approach, predicts novel phenotypes of the *Shewanella* species by quantification of molecular functions in their genomes and by following pair-wise comparison of enriched molecular functions across the species (Fig. 1b). The approach is implemented using network analysis tools. Using the top-down approach, we are able to identify specific genomic clusters related to cold tolerance of the *Shewanella* species, to predict novel cellular mechanisms underlying the bacterial cold tolerance, and to link this trait to salt tolerance of the species. By using the second approach, we reveal novel, environmentally important phenotypes in the species.

Materials and methods

Bacterial strains and culture conditions for testing *Shewanella* species cold and salt tolerance The following 15 *Shewanella* strains were used in testing of the phenotypic traits: *Shewanella oneidensis* MR-1; *Shewanella amazonensis* SB2B; *Shewanella putrefaciens* CN-32; *Shewanella* sp. PV-4; *Shewanella denitrificans* OS217; *Shewanella baltica* OS155;

Fig. 1 Top-down (a) and bottom-up (b) computational frameworks for relating phenotypic and genomic characteristics of the *Shewanella* species based on quantification of molecular functions in the bacterial genomes

(a) **Step 1.** Quantify the phenotypic trait by assigning a score for each species



Step 2. Calculate enrichments of each genome with protein families

Protein family	S. sp. MR-4	S. sp. MR-7	S. sp. ANA-3	S. sp. W3-18-1	S. oneidensis MR-1	S. litchica PV-4	S. ballica OS155
Pfam 1	18	17	19	17	1	13	15
Pfam 2	41	41	43	45	49	43	44
Pfam 3	4	4	4	4	4	4	4
Pfam 4	5	5	5	4	5	5	5
Pfam 5	3	3	4	3	5	3	3
Pfam 6	3	3	3	3	3	3	3

Step 3. Correlate the score and protein family enrichments among species



Step 4. Analyze phenotype specific protein families in terms of genes and gene clusters

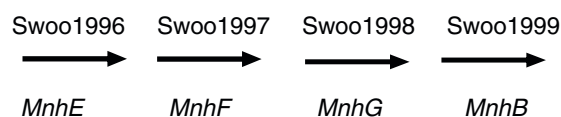
Pfam Model

MNHE

MrpF_PhaF

PhaG_MnhG_YufB

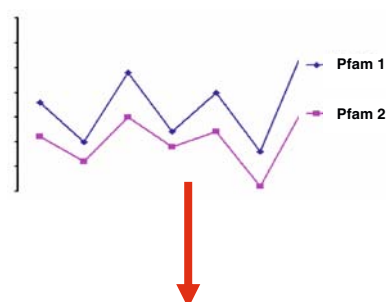
MnhB



(b) **Step 1.** Calculate enrichments of each genome with protein families

Protein family	S. sp. MR-4	S. sp. MR-7	S. sp. ANA-3	S. sp. W3-18-1	S. oneidensis MR-1	S. litchica PV-4	S. ballica OS155
Pfam 1	18	17	19	17	1	13	15
Pfam 2	41	41	43	45	49	43	44
Pfam 3	4	4	4	4	4	4	4
Pfam 4	5	5	5	4	5	5	5
Pfam 5	3	3	4	3	5	3	3
Pfam 6	3	3	3	3	3	3	3

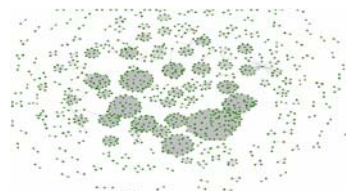
Step 2. Calculate pair wise affinity between domain enrichments among species



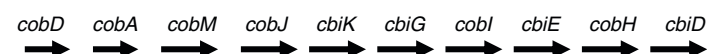
Step 4. Apply a network based clustering to find relating molecular functions representing a phenotype

Cluster	Pfam Model
28	CobU
28	CobS
28	CbiC
28	CbiD
28	CbiG

Step 3. Select highly correlating domain pairs and produce a domain affinity map



Step 5. Analyze phenotype specific domains in terms of genes and gene clusters



Shewanella baltica OS195; *Shewanella woodyi* ATCC 51908; *Shewanella pealeana* ATCC 700345; *Shewanella putrefaciens* 200; *Shewanella* sp. ANA-3; *Shewanella* sp. MR-4; *Shewanella* sp. MR-7; *Shewanella* sp. W3-18-1; and *Shewanella frigidimarina* NCIMB400.

Cells were grown aerobically on *Shewanella* Federation (SF) medium (pH 7.0) containing 3 mM PIPES, 2.8 mM NH_4Cl , 0.4 mM NaH_2PO_4 , 3 mM NaCl, 0.1 mM KCl, 1 μM Na_2SeO_4 , 10 μM L-arginine hydrochloride, 13 μM L-glutamate, 19 μM L-serine, and 10 ml each of 100 \times Wolfe's vitamin and 100 \times mineral solutions prepared as described previously (Kieft et al. 1999) (Supplementary Protocol). Then, 10 mM of lactate was added as the carbon and energy source. Additional sodium chloride (2.5%) was added to the medium of *S. woodyi*. For *S. denitrificans* OS217, experiments were conducted with Luria–Bertani (LB) broth because of its inability to grow on SF medium supplemented with lactate. Starter cultures were initiated by overnight growth in 5 ml of LB or Marine broth 2216 (*S. pealeana* and *S. woodyi* only) on a VWR shaker (speed=125 rpm) at 22°C. Cells were collected by centrifugation at 4,000 rpm for 10 min, washed twice in SF medium lacking lactate, and resuspended in 1 ml of SF medium to inoculate 4 ml SF medium in 10-ml tubes. Temperatures tested ranged from 0 to 55°C (4, 12, 18, 24, 30, 37, 42, and 55°C), and the salinities tested were 0, 0.5%, 1.0%, 2.5%, 5.0%, and 10% NaCl. At least two cultures of each species were grown at each temperature and salinity tested (Supplementary Tables 1S and 5S). Growth was monitored by measuring absorbance at 600 nm with a visible light spectrophotometer (Spec 20). Optical density (OD_{660}) readings were recorded throughout the growth period.

Quantification of phenotypic traits In order to capture the most distinct features of the phenotypes and diminish the effect of the culturing conditions, the traits were scored. The effect of temperature on growth was characterized by the cold tolerance score (CTS), and the effect of salinity on growth was estimated by the salt tolerance score (STS).

To calculate the cold tolerance score we first determined the Growth Scores (GS) for each characterized temperature (T_j) from the growth profiles, namely, values of OD_{660} (OD_i) against time points (t_i). GS values were calculated as the area under the growth profile normalized to 100 h of the growth: $\text{GS} = (1/2) \times (\sum (\text{OD}_{i+1} + \text{OD}_i) \times (t_{i+1} - t_i)) \times 100/t_n$ where $i=0, \dots, n-1$, n is the number of time points, and t_i is the time values during the bacterial growth when OD was monitored. The cold tolerance scores (CTS) were then calculated for each species as the ratio of GS at 4°C to that at 24°C (optimal temperature) and expressed as percentage; thus, the greater the CTS value, the greater the cold tolerance of the species.

The calculation of the salt tolerance score was also based on comparison of the GS values calculated as described above. The STS was calculated as the ratio of GS at 5% of NaCl versus growth without NaCl; thus, the greater the score, the greater salt tolerance or salt requirement of the species.

Quantification of molecular functions and their mapping to gene annotations Nineteen *Shewanella* strains were analyzed either as complete genomes (*S. amazonensis*, *S. baltica* strains OS155 and OS195, *S. denitrificans*, *S. frigidimarina*, *S. loihica*, *S. oneidensis*, *S. pealeana*, *S. woodyi*, *Shewanella* sp. strains ANA-3, MR-4, MR-7, and W3-18-1) or as draft genomes with gene annotations (*S. baltica* strains OS185 and OS223, *S. putrefaciens* strains 200 and CN-32, *S. benthica*, and *S. sediminis*). The number of protein domains in each genome was used as a quantitative parameter to characterize the importance of the molecular function of the protein domain. A list of Pfam domains and gene annotations were downloaded from the US Department of Energy Joint Genome Institute (JGI) <http://www.jgi.doe.gov/>. Since *S. oneidensis* and *S. benthica* KT99 were not sequenced by JGI, their sequences were downloaded from the J. Craig Venter Institute (<http://www.jcvi.org>) and annotated using the JGI pipeline. Briefly, genes were modeled using software packages Critica 1.05 and Glimmer 3.1 (Badger 1999; Delcher et al. 1999). A combination of BLAST (1e–5 cutoff), HMMER (1e–10 cutoff), and PRIAM (1e–30 cutoff) were used to assign functions to the sequences. In addition, the revised gene/protein set was searched against the KEGG GENES and InterPro databases. The number and locations of the protein domains were mapped to the genes in each genome using SQL queries and MS Access software.

Computational framework for relating phenotypic and genomic characteristics The top-down approach for relating phenotypic and genomic characteristics (Fig. 1a) is based on quantification of cold and salt tolerance from growth curves of 15 species as described above. Pearson correlation coefficients (R) are calculated between the CTS and STS and the domain enrichments among the species. To find a threshold for the coefficient, the correlations with the cold tolerance score were calculated by permutation of the domain numbers among species. The permutations gave a distribution of the correlations that was close to normal with the average value of the coefficient –0.0012 and the standard deviation 0.27 indicating that ~90% of random correlations are expected to be less than 0.44. This value was used for initial selection of the domains correlated with the cold tolerance score. The domains were further analyzed using the synteny evidence as described below.

The bottom-up approach (Fig. 1b) does not involve a quantification of bacterial phenotypes and is solely based on quantification of molecular functions in the genomes of 19 sequenced *Shewanella* strains. Molecular functions were quantified by the number of Pfam domains or domain enrichments as described above. Affinities between pairs of domains were estimated using the Pearson correlation. Correlated domain pairs (the Pearson coefficient is less than a threshold) were selected to produce an affinity graph, where the Pfam domains are the nodes connected by the edges if they are correlated. The threshold for correlating domains was found by testing different R values (0.80, 0.85, 0.90, and 0.99). WebR environment was used for the statistical calculations and VisANT 3.04 system for visualization of the networks (Hu et al. 2005). To find subsets of pair-wise correlating domains, which potentially reveal relating molecular functions in the genome, we applied Markov clustering algorithm (Enright et al. 2002). This algorithm is remarkably robust to graph alterations and shows superiority for the extraction of clusters from interaction networks (Brohee and van Helden 2006). The inflation value was set to 1.8 as recommended by the software. The automatic cluster annotation was made by a Perl script. The script assigned a name to each cluster comprised of “+”/“−” followed by the concatenated nicknames of the microorganisms. If the number of microorganisms with overrepresented domains was less or equal 10, then the cluster name had “+” and concatenated nicknames of the microorganisms with overrepresented number of domains in the cluster. Otherwise the cluster name had “−” and concatenated nicknames of the microorganisms with underrepresented number of domains (Supplementary Table 6S). Synteny evidence for identified clusters was extracted considering the conserved gene order in the organisms having an overrepresented number of domains as described below.

Synteny evidence A program written in R (Ihaka and Gentleman 1996) was used to simulate the relationship between (1) the probability for two and more neighboring genes to cluster together and (2) the number of randomly sampled genes. The strand orientation of the neighboring genes was not taken into account during simulation. The probability was estimated empirically by repeating the random sampling of m numbers, where m simulated the cluster size changing from 2 to 250, from the ordered array of numbers from 1 to 4,000, where 4,000 indicated the average number of coding sequences in the *Shewanella* genome. The probability was calculated as a ratio of positive results from 100,000 repeating samplings for each cluster size. Results indicate a low probability of finding two adjacent genes in the cluster by chance, if the cluster

size is less than 10. The probability significantly decreases when the cluster size decreases or when the number of identified neighbors increases. For small clusters (less than five genes), even two neighbors with a gap have a small chance to be found by the random gene selection. Based on the empirical probability simulations we decided to search three types of synteny in the identified clusters as additional evidence that the cluster assembly does not occur by chance alone and is likely driven by a common phenotype. The synteny types included (1) two neighboring genes (N), (2) two neighboring genes with a gap (G), and (3) a synteny loci (L) with three and more neighboring genes. After mapping domains to genes, we used a Perl script to identify and characterize the syntenies in each cluster (Supplementary Table 7S). This information was used to select clusters for validation based on known phenotypes in the MR1 strain (Table 1), which is the most studied organism, and for discovery of new phenotypes in recently sequenced species (Table 2).

Results and discussion

Revealing molecular functions involved in bacterial adaptation to cold temperatures by the top-down approach

Cold tolerance phenotype of *Shewanella* species Although all tested *Shewanella* species have similar optimal growth temperatures with an average value about 24°C and must be classified as mesophilic (Vieille et al. 2001), they show significant differences in their ability to support growth when the temperature dropped to 4–12°C (Supplementary Tables 1S, Supplementary Fig. 2S). Some species show only small changes in growth (accumulated biomass) at the lower temperatures indicating a striking adaptability to the cold conditions. Other species are less tolerant and show a significant impairment of growth. To characterize the phenotypic difference among species, we compared changes in the biomass accumulated by each species at 4°C versus 24°C. By using this ratio, we made the parameter less dependent on the absolute value of the biomass or on the organism’s ability to utilize specific resources and more dependent on its ability to respond to lower temperatures. A higher score reflects a greater the cold tolerance. We referred to the calculated parameter as the cold tolerance score and to the phenotypic trait as cold tolerance. Results of the cold tolerance score calculations (Fig. 2a) indicated significant differences in the adaptability of the *Shewanella* strains to low temperature. *S. pealeana*, a strain isolated from the nidamental gland of the squid *Loligo pealei*, showed the greatest cold tolerance. A significantly lower cold tolerance was observed for *S. amazonensis* SB2B and *S. loihica* PV-4,

Table 1 Domain clusters representing known phenotypes in *S. oneidensis* MR-1

Cluster IDs	Domains comprised the clusters (Pfam models)	Synteny evidence	Related Genomic cluster (locus tags or gene names)	Known Phenotypes	Ref.	Flanking transposases	Other species inferred from the cluster annotation that likely express the same phenotype
22, 172 ^a	Complex1_24kDa; Complex1_30kDa; Complex1_49kDa; Complex1_51K; NADHdh; Oxidored_q1; Oxidored_q1_N; Oxidored_q3; Oxidored_q4; Oxidored_q1; Oxidored_q1_N; Oxidored_q2	SO_1011; SO_1013; SO_1015; SO_1017; SO_1018; SO_1019; SO_1021; SO_1009– SO_1012	<i>nqr-2</i> operon: <i>nuo</i> ABCDIFGHIJKLMN	Na(+)- transporting NADH: ubiquinone oxidoreductase <i>nqr-2</i> operon	(Mrazek et al. 2006)	SO_1008 is ISSod20 transposase, TnpA_ISSod20	<i>S. benthica</i> and <i>S. woodyi</i>
5, 68 ^a	DNA_circ_N; GP46; Phage_Mu_Gp45; Phage_Mu_P; DUF935; Phage_Mu_F; DUF1320; DUF1018	SO_0667; SO_0668; SO_2680; SO_2681; SO_2698– SO_2701	Prophage loci SO_2667–SO_2701	Prophage MuSo2	(Chourey et al. 2006; Qiu et al. 2006)		
105	BATS; Fe_hyd_SSU; Fe_hyd_lg_C; Radical_SAM	SO_3920– SO3923	Formate dehydrogenase Fdh, periplasmic [Fe–Fe] hydrogenase HydAB, and assembly proteins HydGXEF	Hydrogen metabolism	(Meshulam-Simon et al. 2007)		<i>Shewanella</i> spp. MR4 and ANA3
150	FdhE; SelA	SO_0104– SO_0105	Formate dehydrogenase fdnGHIE and L-seryl- tRNA selenium transferases SelA, MetY, MdeA	Formate utilization	(Romine et al. 2004)		<i>Shewanella</i> spp. MR4, MR7, ANA3, <i>S. baltica</i> strains OS223, OS195, and OS185, and <i>S. pealeana</i>

^a The phenotype was represented by two clusters

strains isolated from the Amazon River delta and the microbial mat at a hydrothermal in the Pacific Ocean, respectively. It is possible that these two latter strains more frequently encounter higher temperatures in their environments and are therefore less adapted to growth in the cold. The optimal growth temperatures at 35°C (*S. amazonensis* SB2B), 25°C (*S. pealeana*), and 18°C (*S. loihica* PV-4) did not reflect their cold tolerance. There also appeared not to be a correlation between phylogenetic distance and cold tolerances reflected by the difference observed for the two *S. putrefaciens* strains, CN-32 and 200.

Selection of Pfam domains related to *Shewanella* cold tolerance

To find Pfam domains related to the cold tolerance, we calculated correlations between the number of Pfam

domains and the cold tolerance score using the Pearson coefficient (Supplementary Table 2S, Fig. 2b). The threshold for selecting significant correlations was estimated by permutating the number of domains among species and calculating correlations of the permuted values with the cold tolerance score as described in “Materials and methods”. Although the selected domains are likely important, either directly or indirectly, for *Shewanella* growth at low (<12°C) temperatures, they also may contain false positive correlations. A typical approach to remove false positive correlations and to estimate their rate is to use a benchmark, or a set of known Pfam domains related to the studied phenotype. When studying complex phenotypes, it is difficult to establish a benchmark. There were no previous attempts to quantify cold tolerances of the species, and the biological mechanisms underlying the cold response are largely unknown and may also differ between psychrophiles and mesophiles. To solve

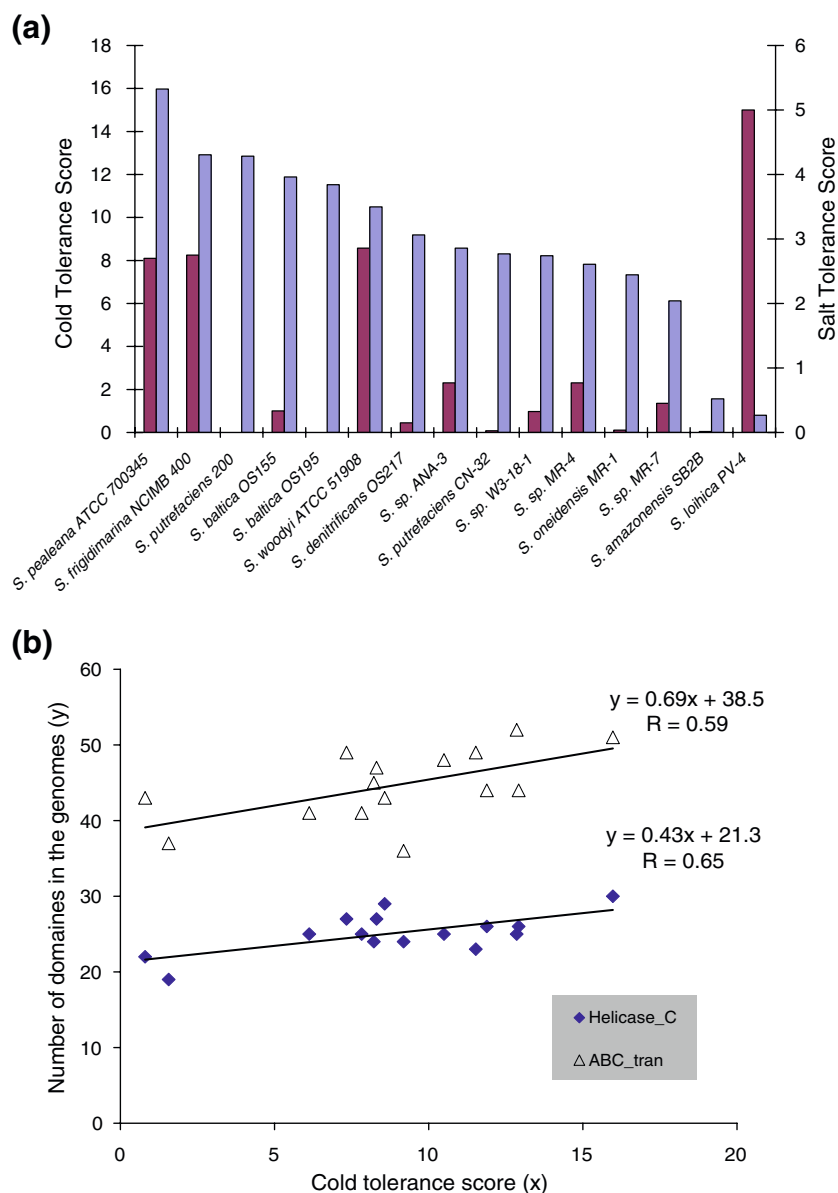
Table 2 Phenotypes of the newly sequenced *Shewanella* species predicted by the bottom-up approach and their characteristics inferred from the identified genomic clusters (see Supplementary Table 8S for more details)

Predicted phenotypes	Species	Phenotype characteristics predicted from the genomic cluster content
Degradation of aromatic compounds by an aerobic hybrid pathway	<i>S. woodyi</i>	The cluster is similar to the <i>paa</i> gene cluster in <i>Pseudomonas</i> sp. (Ferrandez et al. 1998; Nogales et al. 2007). The aerobic degradation involves a novel “hybrid” principle of the catabolism of phenylacetic acids with the involvement of CoA derivatives. It resembles the conventional strategy of anaerobic degradation of aromatic compounds
Bioluminescence (known phenotype)	<i>S. woodyi</i>	The cluster represents the <i>lux</i> operon responsible for luminescence (Nelson et al. 2007). The order of genes in the <i>S. woodyi</i> operon is different from the other known marine bioluminescent bacteria
Ethanolamine utilization	<i>S. benthica</i> (draft genome)	The cluster is similar to the <i>eut</i> operon in <i>Salmonella enterica</i> (Brinsmade et al. 2005)
Anaerobic biosynthesis of vitamin B ₁₂	<i>S. benthica</i> and <i>S. sediminis</i>	Products encoded by the clusters are similar to enzymes involved in the cobalamin synthesis under anaerobic conditions (Roth et al. 1993). In <i>S. benthica</i> , the aerobic and anaerobic pathways of B ₁₂ biosynthesis coexist. The anaerobic route is likely employed when ethanolamine is utilized considering co-localization of the <i>eut</i> and <i>cob</i> loci and the requirement of B ₁₂ for the growth on ethanolamine
Propanediol degradation	<i>S. putrefaciens</i> CN32 and <i>Shewanella</i> sp. W3181	The genomic clusters are similar to the <i>pdu</i> operon in <i>Salmonella</i> (Bobik et al. 1999). The propanediol degradation in <i>S. putrefaciens</i> CN32 and <i>Shewanella</i> sp. W3181 presumably occurs only aerobically, because propanediol is utilized in a coenzyme B ₁₂ -dependent fashion and neither organism has the phenotype of anaerobic biosynthesis of B ₁₂ identified in <i>S. benthica</i> and <i>S. sediminis</i>
Arsenic resistance	<i>S. putrefaciens</i> strains CN-32 and 200, <i>Shewanella</i> sp. W3-18-1 and ANA-3	The clusters include <i>arsD</i> and <i>arsA</i> genes that confer an increased rate of extrusion and resistance to arsenite (Lin et al. 2007). A recent study of the <i>ars</i> operon in strain ANA-3 provided evidence of the operon function under either aerobic or anaerobic conditions (Saltikov et al. 2003)
Copper resistance	<i>S. benthica</i> KT99, <i>S. denitrificans</i> OS217, <i>S. frigidimarina</i> NCIMB 400, <i>Shewanella</i> sp. ANA-3, <i>S. putrefaciens</i> CN-32	The clusters are similar to the <i>cop</i> operon in <i>Pseudomonas syringae</i> (Maher et al. 2006). In all five genomes, the first gene in the cluster is a conserved hypothetical protein (DUF411), which likely encodes the copper-responsive repressor CopR that is often transcribed as a part of the <i>cop</i> operon (Magnani et al. 2008)
Mating pair formation to transfer DNA and other macromolecules	<i>S. baltica</i> strains OS223, OS195 and OS185, <i>Shewanella</i> sp. W3-18-1 and ANA-3 (surface-exposed sex pilus only)	The cluster encodes products of the type IV secretion system (Lawley et al. 2003). Interestingly, this system is implemented in the <i>Shewanella</i> species with different levels of complexity. It is clearly indicated by the number of Pfam domains
Uptake and release of H ₂	Absent from <i>S. denitrificans</i> and <i>S. woodyi</i> only	The genomic clusters are similar to the periplasmic [Ni–Fe] hydrogenase <i>hyaB</i> (Armstrong and Albracht 2005). The absence of the hydrogenase in <i>S. denitrificans</i> and <i>S. woodyi</i> reveals that these two species may be deficient in their metabolic capabilities compared to the other sequenced <i>Shewanella</i> species

the benchmark problem, we used the conserved gene neighborhood of the identified domains among species as a filter. There is very low probability that a random sampling of a small number of genes (less than 10) into a cluster will result in two to three consecutive genes. According to our empirical

modeling (see “Materials and methods” and Fig. 3), an alternative hypothesis (three consecutive genes will not cluster together as a result of random sampling of 30 genes out of 4,000 genes in the genome) has a *p* value less than 0.0015. Thus, finding three to four consecutive genes in the

Fig. 2 Cold (blue) and Salt (violet) tolerance scores for tested *Shewanella* strains and two examples (a) of correlation between the cold tolerance score among the strains and the number of domains that represent helicase conserved C-terminal domain (Helicase_C) and ABC transporter (ABC_tran) (b)

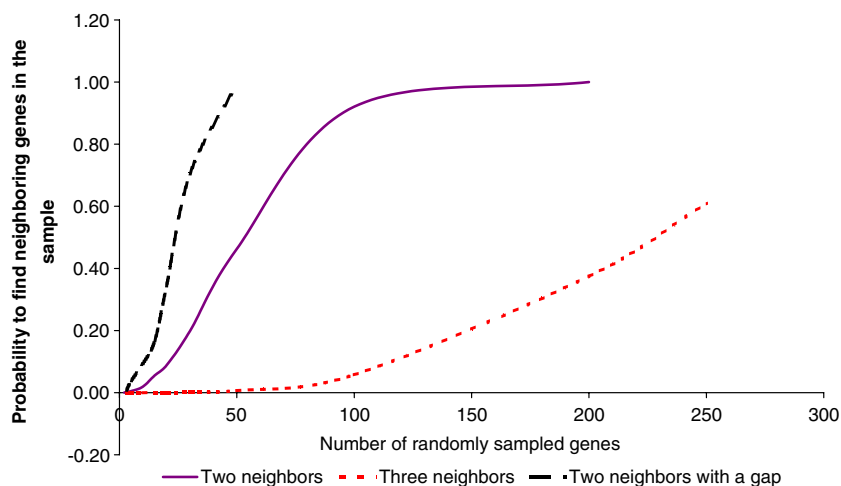


domain cluster provides very stringent criteria for revealing domains that correlate with the cold tolerance score.

Information on conserved gene neighborhoods, also referred to as synteny, was obtained by mapping Pfam domains to genes in the *S. pealeana* genome (Supplementary Tables 3S and 4S) as this organism has the greatest cold tolerance of the tested species. The search indicated a set of four domains in *S. pealeana*, *S. woodyi*, and *S. frigidimarina* that mapped to four consecutive genes representing hydrophobic subunits of an Na^+/H^+ antiporter. This gene cluster was followed by six genes encoding subunits of a respiratory NADH-ubiquinone oxidoreductase (Fig. 4). Both of these clusters likely belong to a single operon, considering the

very short intergenic regions and the overlap of coding sequences in the organisms. The presence of this operon in the three most cold-tolerant species suggests that the Na^+ -based pump is involved in cold tolerance. Indeed, the use of sodium ions instead of protons for generation of the motive force is a known microbial strategy in response to changes in the fluidity of the cytoplasmic membrane (Chattopadhyay 2006; Konings 2006). Many marine and pathogenic bacteria couple the Na^+ pump to the respiratory chain (Kogure 1998; Hase et al. 2001; Dibrov et al. 2004). The gene clusters we have identified in cold-tolerant *Shewanella* species are similar to the Na^+/H^+ antiporter clusters found in the halotolerant bacterium *Staphylococcus aureus* (Hiramatsu et

Fig. 3 The empirically calculated probability for finding neighboring genes among a randomly sampled cluster of genes in the genome comprised 4,000 genes. The probability was calculated by random sampling genes in the clusters



al. 1998). Thus, the cold tolerance of *Shewanella* species may also entail their salt tolerance or possibly a sodium requirement for growth.

Phenotypic correlation between cold and salt tolerance of the species Although growing evidence associates bacterial cold tolerance with adaptation to other stresses (Chattopadhyay 2006), a link between cold and salt tolerance and an association with the Na^+/H^+ antiporter gene clusters has not been previously reported. To further evaluate this phenotypic association, we experimentally tested the effect of different NaCl concentrations on the growth of the *Shewanella* species (Supplementary

Table 5S). Salt tolerance scores were calculated for each strain as the ratio of OD_{660} accumulated after 1 week of incubation of the strain with 5% of NaCl versus culturing without NaCl. Thus, the greater the value of the score, the greater salt tolerance of the species and its salt requirement. Figure 2a shows a comparison of the species according to their salt tolerance scores, and it indicates that the three most cold-tolerant species, *S. pealeana*, *S. woodyi*, and *S. frigidimarina*, with a predicted Na-based respiration have very similar salt tolerances. Their salt tolerance scores are significantly greater (more than three times) than those of the rest of the tested *Shewanella* species. The only exception is *S. loihica* PV-4. The salt

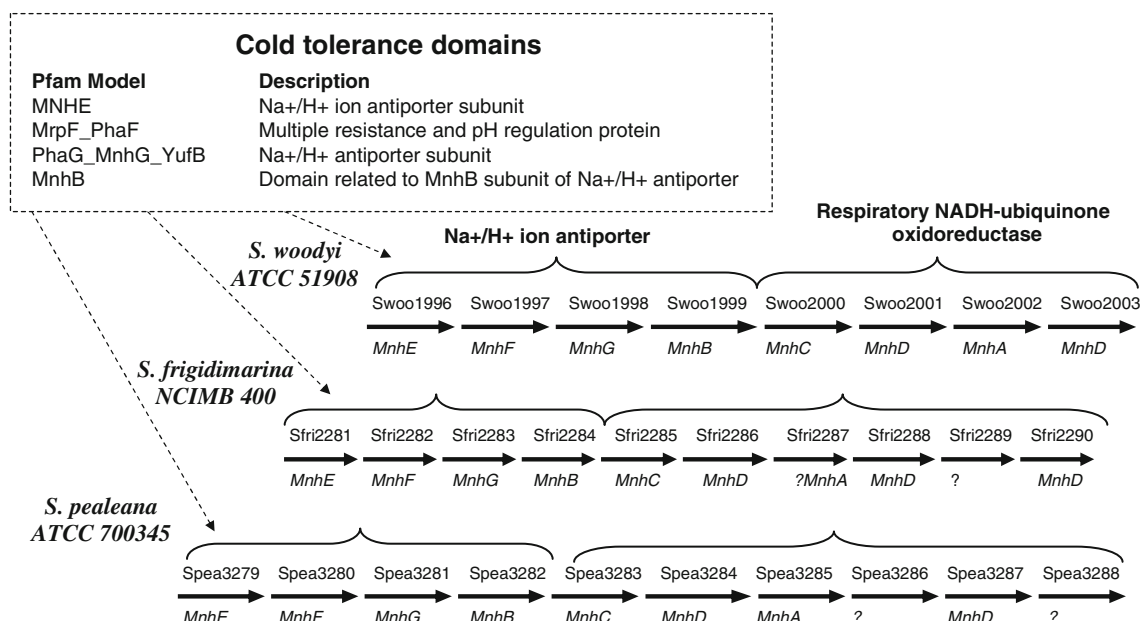


Fig. 4 Schematic map of the gene regions in *Shewanella* sp. with the cold tolerance domains representing Na^+/H^+ antiporter and NADH-ubiquinone oxidoreductase

tolerance score of *S. loihica* PV-4 is two times greater than the scores of the *Shewanella* species with the Na^+/H^+ antiporter. This suggests that the genetic mechanism underlying the exceptional salt tolerance and requirement of *S. loihica* PV-4 differs from that of *S. pealeana*, *S. woodyi*, and *S. frigidimarina* NCIMB 400.

The top-down approach identified many known protein functions involved in bacterial cold tolerance

The synteny analysis allowed us to establish a more conservative threshold for correlation of the cold tolerance score with number of Pfam domains. We revealed 109 domains that likely relate to cold tolerance (Supplementary Tables 2S and 3S). Indeed, 24 of these domains represent known mechanisms of cold tolerance including (1) maintenance of the membrane fluidity (Phadtare 2004) by introduction of double bonds into fatty acids via desaturases (FA_hydroxylase), (2) enhanced translation and transcription (Nakaminami et al. 2006; Wilson 2007) indicated by the presence of unique tRNA synthetases and chaperons (DALR_2, TruB-C_2, tRNA-synt_1g, Elong-fact-P_C, HAS-barrel), (3) increased proteolysis (Peptidase_C39, Peptidase_U32, Leuk-A4-hydro_C, Creatinase_N), and (4) transport of ions (ABC_tran, Aa_trans, Peripla_BP_2, OpuAC, Sec_GG, Glyphos_transf, Multi_Drug_Res, DcuA_DcuB, MotA_ExbB, UAA, MnhB, MrpF_PhaF, MNHE, PhaG_MnhG_YufB) (Erdal et al. 2003; Susana et al. 2004; Vezzi et al. 2005; Auman et al. 2006; Chattopadhyay 2006; Konings 2006). Several of the domains (OmpA_membrane, DHHA2, DHH, Aldedh, Oxidored_FMN, Elong-fact-P_C, Surf_Ag_VNR) are among the genes and proteins activated by cold shock in *Shewanella piezotolerans* WP3 and *Shewanella livingstonensis* Ac10 (Venkateswaran et al. 1999; Kawamoto et al. 2007).

Revealing novel genomic loci of bacterial cold tolerance by the proposed approach

We identified 10 *S. pealeana* loci with three or more consecutive genes that have a very high probability of being linked to cold tolerance (Supplementary Table 4S). These loci are believed to represent novel mechanisms of cold tolerance. In addition to the Na^+/H^+ antiporter considered above, one loci (Spea_0901–Spea_0922) includes genes encoding two helicases, a thiol-specific antioxidant, a valyl-tRNA synthetase, and two peptidases. Another cold tolerance locus (Spea_3491–Spea_3502) includes two chemotaxis sensory transducer proteins, a cysteine exporter, a helicase, and several hypothetical proteins. Similarity searches show that the chemotaxis sensory transducer proteins (Spea_3491 and Spea_3492) have close homologs in the barophilic and psychrophilic

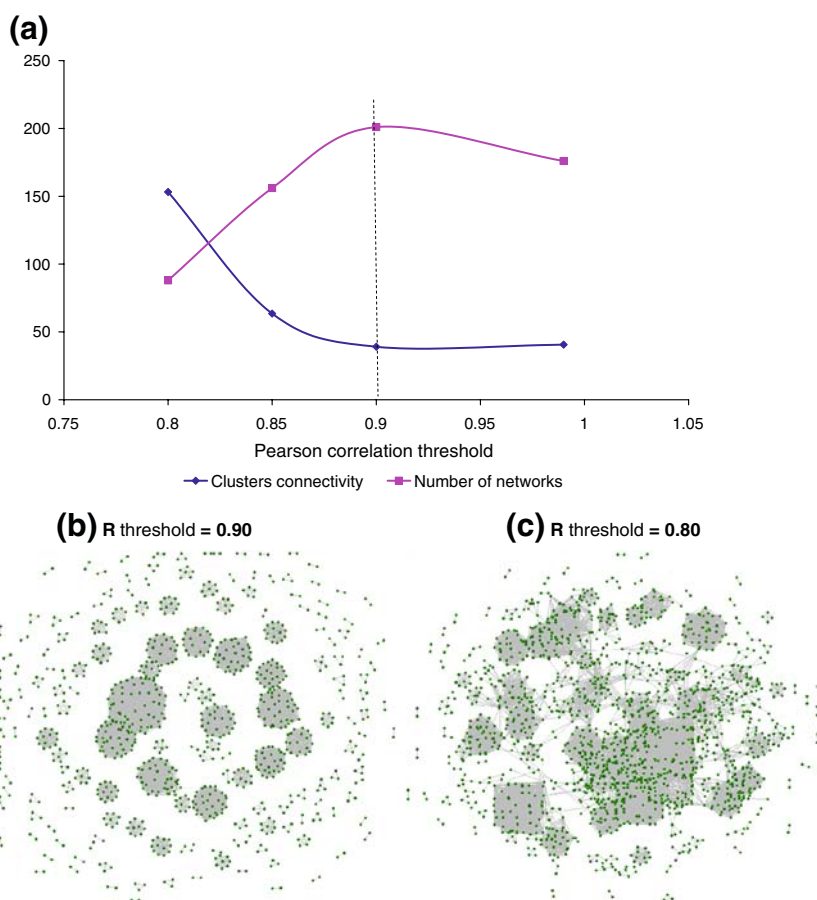
bacterium *Photobacterium profundum* SS9 (Vezzi et al. 2005) and that the cysteine exporter is homologous to cysteine transporters (Ping_1298 and Ping_3454) of the psychrophilic bacterium *Psychromonas ingrahamii* isolated from Arctic polar sea ice (Auman et al. 2006). The presence of proteins with DEAD/DEAH box helicase domains in three of the identified loci strongly links RNA metabolism to resistance to low temperature. It is known that these helicases loosens RNA secondary structure and promotes folding of RNA (Polissi et al. 2003). This may be important at lower temperatures, because they stabilize the RNA secondary structures, and this stabilization is a major detrimental physiological effect of temperature downshift.

Predicting phenotypes of *Shewanella* species from comparative analysis of molecular functions using the bottom-up approach

The approach (Fig. 1b) was applied to 19 sequenced *Shewanella* strains. Similar to the top-down analysis, the selection of clusters related to phenotypes was initially based on the correlation coefficient threshold and then further augmented by synteny information inside each cluster by mapping domains to genes and searching for neighboring genes and loci.

The analysis included three main computational steps: (1) calculate a profile for each domain comprised of numbers of the domain in the tested genomes (Supplementary Table 6S), (2) calculate Pearson correlations for each pair of the domain profiles, and (3) find disconnected networks by clustering the correlated profiles according to their pair-wise correlations using a threshold. To find an optimal threshold for the correlation, the clustering step was repeated using different thresholds for the correlation coefficient. The ultimate goal of repeating the calculation was to find the correlation coefficient that maximizes the number of the identified networks and minimizes the networks/clusters connectivity. By maximizing the number of networks, we increase the number of phenotypes we can reveal, and by minimizing the connectivity between networks we increase specificity of domain attribution to each network. The selected threshold, therefore, provided a reasonable tradeoff between specificity and sensitivity of the phenotype identification. Results of the threshold selection are presented in Fig. 5 and indicate the optimal value for the correlation coefficient $R=0.90$. The affinity map includes 201 disconnected networks comprised of two and more domains, which after the Markov clustering separated into 217 clusters. The clusters and networks mainly represent sets of domains specific to one or several *Shewanella* species. They were annotated automatically (with manual validation) by nicknames of the organisms having overrepresented or underrepresented numbers of

Fig. 5 Selection of the domain affinity threshold. The domain affinity is estimated by the Pearson correlation coefficient (R) and different thresholds of the coefficient were tested to visualize the obtained domain affinity maps and to calculate the number of pairs that are greater than the thresholds, the number of networks (disconnected clusters), and the cluster connectivity (the ratio of selected pairs to the number of network). The optimum threshold (*dashed line*) for R was selected to maximize the number of the identified networks and to minimize the cluster connectivity (a). Figures (b) and (c) demonstrate difference between networks based on different thresholds



domains (Supplementary Table 6S). To assign each cluster to a phenotype and to augment validity of the assignment, we extracted synteny information from each cluster by mapping domains to genes (Supplementary Table 7S). We found that 84 of the 217 identified clusters have additional synteny evidence, i.e., two or more adjacent genes in the genome. Thirty-five clusters consisted of loci with three to eight conserved genes. We further analyzed the clusters to validate the bottom-up approach and to discover novel phenotypes in the *Shewanella* strains. The synteny information in the clusters included (1) neighboring genes; (2) neighboring genes with a gap, which is one gene between neighbors; and (3) a genomic locus of three or more neighboring genes.

For validation of the approach, we selected seven clusters with orthologs in *S. oneidensis* MR-1. We reasoned that since *S. oneidensis* MR-1 is an experimentally well-studied environmental bacterium, the clusters are more likely to be linked to a phenotype. Indeed, we find that the clusters can be linked to four known MR1 phenotypes, and that the expression of genes related to each phenotype has the experimental support in the literature (Table 1). In addition to validating the approach, the enrichment of these domains allows us to infer similar phenotypes for the newly sequenced genomes containing these clusters.

Most of the identified clusters did not include *S. oneidensis* MR-1 and represented phenotypes that are novel for other *Shewanella* species. The clustering augmented by synteny information facilitated the linking of clusters to phenotypes and highlighted loci of phenotype related genes in two and more genomes. We selected 15 clusters with different types of synteny evidence for manual evaluation of phenotypes (Supplementary Table 7S) using literature-based information. We find that the selected clusters represent nine phenotypes in *Shewanella* species, including degradation of aromatic compounds by an aerobic hybrid pathway in *S. woodyi*, degradation of ethanolamine by *S. benthica*, and propanediol degradation by *S. putrefaciens* CN32 and *Shewanella* sp. W3-18-1. Arsenic and copper resistance phenotypes were detected for some *Shewanella* species, as well as differing abilities of the species to transfer macromolecules and H_2 . Table 2 and Supplementary Table 8S provide some characteristics of the phenotypes and their specificities discovered from the cluster content of the organisms. A comparison of the clusters with metabolic reconstructions provided by the *Shewanella* Knowledgebase (<http://shewanella-knowledgebase.org:8080/Shewanella/shewcyc.jsp>) indicated that none of the phenotype could be predicted based on the enzymatic annotation.

Applications and limitations of the proposed approaches

The results of our study indicate that quantifying complex phenotypes and combining statistical and synteny criteria are valuable novel computational tools to study the molecular mechanisms behind the adaptation of an organism's physiology to its environment. As a rule, the adaptation has a continuing nature and cannot be characterized simply as present or absent. The quantification of the molecular function in terms of the number of domains instead of a binary characterization (presence/absence) is equally important. Although, in many cases, domains that are unique to an organism indicate a specific phenotypic trait, the enriched domains may reveal additional molecular functions that, in combination with the organism-specific domains, more accurately map domain clusters to gene loci underlying the phenotype. One example in Table 2 is the identification of two additional gene clusters encoding enzymes involved in anaerobic biosynthesis of B₁₂ in *S. benthica*. This vitamin is an essential cofactor for the utilization of ethanolamine, a specific phenotype of this bacterium. A combination of enriched and specific domains was mapped to a gene cluster in the genome that was attributed to a specific phenotypic trait of the bacterium. The other example is an increased number of the BMC domains relating to carboxysome-like structural proteins that was found in three of the species. These proteins are structural components of bacterial microcompartments used for utilization of ethanolamine in *S. benthica* and propanediol in *S. putrefaciens* CN32 and *Shewanella* sp. W3-18-1. In both cases, the microcompartments are believed to diminish aldehyde toxicity.

A noticeable feature of most genomic loci attributed to phenotypes is the presence of transposases and integrases immediately upstream or downstream of the mapped genes (Table 1, Supplementary Table 8S). This suggests that many phenotypes are acquired by horizontal gene transfer and that this mechanism plays an important role in the innovation and evolution of the *Shewanella* genomes. Known phylogenetic methods for identification gene transfer events look for unusual distributions of genes across organisms. Phylogeny-independent methods look for genes that appear abnormally in their genomic context by examining nucleotide frequencies or codon usage bias. Both methods often produce inconsistent results (Gogarten et al. 2002). The proposed algorithm automatically highlights clusters in a genome that have both abnormal sequences and patchy distribution across the genomes and provides a novel method for collecting evidence of potential gene transfer events.

Our proposed approaches provide valuable tools not only for prediction of phenotypes and revealing genomic content underlying complex physiological characteristics

but also for prediction of orthologs in the genomes, and functions of unknown domains and hypothetical proteins. A genome-scale search for orthologs is usually performed by a reciprocal-best-BLAST-hits approach (Altschul et al. 1990). Such predictions, however, are based solely on sequence similarity between two genes and do not take into consideration the nearby genes or the domain structure of the sequences. The domain clustering and subsequent mapping to genes reveal not only individual orthologs in the genomes but sets of orthologs that are common across organisms and relate to shared biological processes or even phenotypes. The approach automatically integrates genomic content into the analysis, thus improving the specificity of the ortholog predictions across organisms without a sophisticated phylogenetic analysis. The prediction of function for unknown domains and hypothetical proteins is another potential application of the bottom-up approach. A good example is DUF59 (PF01883) a domain of unknown function that is found in the cluster representing phenylacetic acid degradation. DUF59 is found in the missing PaaI subunit of phenylacetate-CoA oxygenase, and the domain is encoded adjacent to the genes for the PaaG and PaaJ subunits of the oxygenase. This finding clearly attributes the molecular function “PaaI subunit of phenylacetate-CoA oxygenase” to DUF59. Another example is DUF411, a domain found in a hypothetical protein of the *Shewanella cop* operons. The protein is likely a regulator of the MerR family based on its sequence similarity to a transcriptional regulator in *Saccharophagus degradans* 2-40. Interestingly, we find that *Shewanella* sp. ANA-3 and *S. putrefaciens* CN-32 have an additional common synteny with the DUF411 domain. A comparison of all proteins with the DUF411 domain in *Shewanella* species (Supplementary Fig. 3S) indicate a higher similarity among the proteins of the same synteny in different species and a lower similarity for proteins belonging to different syntenies within the same genome. This is probably reflective of divergence of paralogs and demonstrates feasibility of the approaches from an evolutionary standpoint (Supplementary Fig. 1S) considered in the introduction in more detail.

Although our proposed approaches provide valuable tools for large-scale linking of phenotypic traits to genetic elements, it should be emphasized that the relationships are predicted computationally. Additional experimental studies are necessary to validate the putative relationships. Our analysis also does not include some important aspects of phenotypes, such as regulation, and is limited to the functions characterized by Pfam domains. Anaerobic respiration in *Shewanella* species, for example, involves various multi-heme cytochromes. These proteins are not characterized by Pfam domains and are therefore excluded from the analysis. The integration of different data types in the computational analysis will make it more robust.

Nonetheless, we believe that the proposed approaches can be used to provide a molecular-level understanding of complex ecophysiological characteristics and predict phenotypes of different organisms that can be evaluated experimentally.

Acknowledgments We are very grateful to Arcady R. Mushegian and other reviewers of the paper for thoughtful suggestions and comments on the study. The sequence data for the *Shewanella* species except *S. oneidensis* MR-1 and *S. benthica* KT99 were produced by the US Department of Energy Joint Genome Institute (<http://www.jgi.doe.gov/>). The sequence data for *S. oneidensis* MR-1 and *S. benthica* KT99 were produced by J. Craig Venter Institute (<http://www.tigr.org/>). This research was supported by the U.S. Department of Energy (DOE) Office of Biological and Environmental Research under the Genomics: GTL Program via the *Shewanella* Federation consortium.

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